

Schedule of Accreditation



Organisation Name	Central Veterinary Research Lab-Pathology & Virology Divisions
Trading As	
INAB Reg No	267T
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Accreditation Standard	ISO 17025 T
Date Initially Awarded	07/09/2010
Scope Classification	Biological and veterinary testing
Services available to the public ¹	No

¹ Refer to document on interpreting INAB Scopes of Accreditation

Sites from which accredited services are delivered	
(the detail of the accredited services delivered at each site are on the Scope of Accreditation)	
Name	Address
1 Head Office	Backwestern Campus, Young's Cross, Celbridge, Kildare, W23 X3PH

Scope of Accreditation

Head Office

Biological and Veterinary Testing

Category: A

Biology/veterinary field - Tests	Test name	Technique	Matrix	Equipment	Std. reference	
804 Detection of bacterial, parasite, viral or fungal antigens using specific antibodies and appropriate techniques - .03 Enzyme immunoassay,	IDEXX ELISA for detection of bovine viral Diarrhoea Virus (types I & II) in bovine serum, ear notches and semen	SOP 11.11	serum, ear notches & semen	IDEXX ELISA kit	IDEXX kit	
	IDEXX ELISA for detection of antibodies to Bovine Leukaemia Virus in bovine serum	SOP 11.13	serum	IDEXX EBL ELISA kit	Manual of Diagnostic Tests and Vaccines for Terrestrial Animals of the OIE - Chapter 2.4.11	
	IDEXX ELISA for detection of antibodies to bovine viral Diarrhoea Virus in bovine serum	SOP 11.12	serum	Herdchek IDEXX BVDV ELISA kit	Manual of Diagnostic Tests and Vaccines for Terrestrial Animals of the OIE - Chapter 2.4.8	
	IDEXX IBRgB ELISA for detection of antibodies to bovine Herpesvirus 1 (BHV1) in bovine serum	SOP 11.10	serum	IDEXX IBR ELISA kit	IDEXX kit Manual of Diagnostic Tests and Vaccines for Terrestrial Animals of the OIE - Chapter 2.4.13	
	IDEXX IBRgE ELISA for detection of antibodies to the gE antigen of bovine Herpesvirus 1 (BHV1) in bovine serum	SOP 11.51	bovine serum	IDEXX ELISA kit	Manual of Diagnostic tests and vaccines for Terrestrial animals - Chapter 3.4.11	
804 Detection of bacterial, parasite, viral or fungal antigens using specific antibodies and appropriate techniques - .07 Neuraminidase inhibition	Neuraminidase Inhibition test to characterise AIV in allantoic fluid	SOP 11.07	allantoic fluid	Antiserum to neuraminidase serotypes	SOP 11.07, based on Vandeusen, et al. (1983):Swaayne, et al. (1998): Pedersen (2007)	
804 Detection of bacterial, parasite, viral or fungal antigens using specific antibodies and appropriate techniques - .09 Agar gel immunodiffusion	Coggins Test for antibodies to EIAV	SOP 11.15	serum	IDVet EIA AGID Antibody test kit	SOP 11.15, based on ANON (2013). Equine Infectious Anaemia: Manual of diagnostic tests and vaccines for terrestrial animals. Chapter 2.5.6, OIE	
805 Detection and/or identification of bacterial, parasite, fungal and viral nucleic acids using	Detection of Avian/Equine Influenza Virus matrix gene by real time RT-	SOP 10.02	tissue, swabs	Real time Quantitative Thermocycler	SOP 10.02, based on Spackman E., et al. (2002). Development of a real time reverse	

appropriate techniques - .04 Nucleic acid amplification tests, in house developed assays	PCR in tissues and swabs				transcriptase PCR assay for type A influenza virus and the avian H5 and H7 haemagglutination subtypes. L Clin. Microbiol. 40, 3256-3260	
	Detection of African Horse Sickness Virus by real time RT-PCR in blood and tissue samples	SOP 10.01	blood & tissue	Real time Quantitative Thermocycler	SOP 10.01, based on Aguero, M., et al. (2008). Real-time fluorogenic reverse transcription polymerase chain reaction assay for detection of African horse sickness virus. J Vet Diagn Invest. 20(3): 325-8	
	Detection of African Swine Fever Virus by real time RT-PCR in blood and tissue samples	SOP 10.05	blood & tissue	Real time Quantitative Thermocycler	SOP 10.05 based on King, D., et al. (2003). Development of a Taqman PCR assay with internal amplification control for the detection of African Swine fever virus. Journal of Virological Methods 107, 53-61	
	Detection of Avian/Influenza virus H7 gene by real time RT-PCR in tissues and swabs	SOP 10.04	tissues & swabs	Real time Quantitative Thermocycler	SOP 10.04, based on Spackman, E., et al. (2002). Development of a real time reverse transcriptase PCR for type A influenza virus and the avian H5 and H7 shaemagglutination subtypes. J. Clin. Microbiol. 40: 3256-3260	
	Detection of Avian/Influenza Virus H5 gene by real time RT-PCR in tissues and swabs	SOP 10.03	tissues & swabs	Real time Quantitative Thermocycler	SOP 10.03 based on Spackman E., et al. (2002). Development of a real time reverse transcriptase PCR assay for type A influenza virus and the avian H5 and H7 haemagglutination subtypes. L Clin. Microbiol. 40, 3256-3260	
	Detection of Blue Tongue Virus genome segment 10 by real time RT-PCR	SOP 10.43	blood	Real time Quantitative Thermocycler	SOP 10.43, based on Wernike, K., Hoffman, B. and Beer, M. (2015). Simultaneous detection of five notifiable viral diseases of cattle by single-tube multiplex real-time RT-PCR. Journal of Virological Methods 217, 28-35	
	Detection of Bluetongue Virus serotypes 1-24 by real time RT-PCR in blood samples	SOP 10.07	blood	Real time Quantitative Thermocycler	SOP 10.07 based on Shaw, A.E., et al. (2007). Development and intial evaluation of a real time PCR assay to detect bluetongue virus genome segment 1. Journal of Virological Methods 145, 115-126	

Detection of Classical Swine Fever Virus by real time RT-PCR in blood samples	SOP 10.09	blood	Real time Quantitative Thermocycler	SOP 10.09, based on McGolrick, A., et al. (1999). Closed one tube reverse transcription nested polymerase chain reaction for the detection of pestiviral RNA with fluorescent probes. Journal of Virological Mehods 79, 85-95
Detection of WNV RNA by Real time RT-PCR	SOP 10.16	tissue	Real time Quantitative Thermocycler	SOP 10.06, based on Linke, S., et al. (2007). Detection of West Nile virus lineages 1 and 2 by real time PCR. Journal of virological methods. 146, 355-388
Multiplex Real Time reverse transcription polymerase chain reaction (MP rtRT-PCR) assay for detection of Porcine Respiratory and Reproductive Syndrome Virus (PRRSV) RNA in porcine serum, tissue and semen.	SOP 10.27	porcine tissue, serum and semen	Real Time Quantitative thermocycler	SOP 10.27 based on Wernike et al (2012). Porcine Reproductive and Respiratory Syndrome Virus: Interlaboratory ring trial to evaluate real-time reverse transcription PCR detection methods. Journal of Veterinary Diagnostic Investigation, 24(5) 855-866.
Real time PCR to detect BHV-1 & BHV-4 in ruminant tissues, semen and swabs	SOP 10.44	tissue, swabs, semen	Real time Quantitative Thermocycler	SOP 10.44, based on Herlekar, D.A., Shashikant, C.S., Gurar, A.A., Jayarao, B.M. (2013). Presence of viral and bacterial organisms in milk and their association with somatic cell counts. Journal of Dairy Science 96, 6336-6346
Real time RT-PCR o detect CEMO in swabs and semen	SOP 10.15	swabs	Real time Quantitative Thermocycler	SOP 10.15, Wakely, P.R., et al. (2006) Development of a real time PCR for the detection of Taylorella equigenitalis directly from genital swabs and discrimination form Taylorella asinigenitalis: Veterinary Microbiology, 118, 247-254
Real time RT-PCR to detect EAV in equine blood, nasal swab, semen and tissue	SOP 10.12	whole blood, plasma, tissue	Real time Quantitative Thermocycler	SOP 10.12, based on Balasuriya, UBR., et al. (2002). Detection of equine arteritis virus by real time taqman reverse transcription PCR assay. J. Virological Methods. 101:21-28
Real time RT-PCR to detect EIAV in equine blood and tissue	SOP 10.10	blood & tissue	Real time Quantitative Thermocycler	SOP 10.10, based on Monney, J., et al. (2006). Equine Infectious Anaemia in Ireland: Characterisation of the virus. Vet. Record 159:570

	Real time RT-PCR to detect FMDV in blood and epithelial tissue	SOP 10.13	serum & epithelial suspensions	Real time Quantitative Thermocycler	SOP 10.13, based on Reid, S., et al. (2002). Detection of all seven serotypes of foot and mouth disease by real time fluorogenic reverse transcription polymerase chain reaction. Journal of Virological Methods 105, 67-80.	
	Real time RT-PCR to detect pestivirus by real time Taqman RT-PCR in blood, epithelial tissue, semen, milk and swabs	SOP 10.08	epithelial suspensions, blood, semen, milk & swabs	Real time Quantitative Thermocycler	SOP 10.08, based on McGoldrick, A., et al (1999). Closed one tube reverse transcription nested polymerase chain reaction for the detection of pestiviral RNA with fluorescent probes. Journal of Virological Methods 79, 85-95	
	Real time RT-PCR to detect SVDV in serum and epithelial tissue	SOP 10.14	epithelial suspensions & serum	Real time Quantitative Thermocycler	SOP 10.14, based on Reid, S., et al. (2004). Evaluation of real-time reverse transcription polymerase chain reaction assays for the detection of swine vesicular disease virus. Journal of Virological Methods 116, 169-176	
	Real-time PCR to detect BHV1 in ruminant tissues, semen and swabs	SOP 10.06	tissue, swabs & semen	Real time Quantitative Thermocycler	SOP 10.06, ANON (2010). Infectious Bovine Rhinotracheitis/Infectious Pustular Vulvovaginitis: Manual of diagnostic tests and vaccines for terrestrial animals. Chapter 2.4.13	
	Real-time reverse transcription polymerase chain reaction (rtRT-PCR) assay for detection of avian avulavirus-1 virus RNA in avian swab, tissue and faecal samples.	SOP 10.31	avian swab, tissue and faecal samples	Real Time Quantitative thermocycler	SOP 10.31 based APMV-1/Avian Avulavirus type 1 screening real time PCR SOP, V1.570 Edition 11, APHA UK (OIE/FAO International reference laboratory for Newcastle Disease.) Issued 10/3/17..	
806 Identification of cultured bacteria and fungi using non-nucleic acid based techniques - .03 Identification of fungi by microscopic morphology	Isolation and Identification of Taylorella equigenitalis	SOP 2.25 based on OIE Terrestrial Manual (2012) chapter 2.5.2 (CEM)	swabs	n/a	OIE Manual (2012) chapter 2.5.2 (CEM)	
808 Detection of antibody response to infection using appropriate techniques - .09 Haemagglutination inhibition	Detection of antibodies to avian Influenza virus H5 and H7 subtypes by Haemagglutination Inhibition test in serum	SOP 11.01	serum	stock antigen, positive/negative control sera	SOP 11.01, based on ANON (2015). Avian Influenza: Manual of diagnostic tests and vaccines for terrestrial animals. Chapter 2.3.4. (2015)OIE	
	Haemagglutination inhibition (HAI) to detect antibodies to	SOP 11.14	serum	red blood cells, stock antigen, control serum	SOP 11.14, based on ANON (2012). Newcastle Disease:	

	avian paramyxovirus-1 in avian sera				Manual of diagnostic tests and vaccines for terrestrial animals. Chapter 2.3.14 (2012) OIE	
808 Detection of antibody response to infection using appropriate techniques - .11 Agar gel immunodiffusion	Detection of antibodies to avian influenza virus matrix gene by agar gel immunodiffusion in serum	SOP 11.02	serum	chick embryos, control serum, antigen	SOP 11.02, based on ANON (2015). Avian Influenza: Manual of diagnostic tests and vaccines for terrestrial animals Chapter 2.3.4 (2015) OIE	
808 Detection of antibody response to infection using appropriate techniques - .14 Serum neutralisation test	Detection of antibodies to Equine Arteritis Virus by virus neutralisation test	SOP 11.16	serum	Control antisera, EAV virus, RK-13 cells, Earle's medium	SOP 11.16, based on ANON (2013) Equine Viral Arteritis : Manual of diagnostic tests and vaccines for terrestrial animals. Chapter 2.5.10	
	Rapid serum agglutination test for Avian mycoplasmosis (M. gallisepticum, M. synoviae, M. meleagridis)	SOP 11.17	serum	Stock antisera	SOP 11.17, based on ANON(2008). Avian Mycoplasmosis: Manual of diagnostic tests and vaccines for terrestrial animals. Chapter 2.3.5, OIE	
810 Culture of virus and other obligate intracellular pathogens using in vivo or in vitro techniques	Virus Isolation and Identification (egg inoculation technique) for avian influenza and avian Paramyxovirus	SOP 11.05	tissue/swabs	embryonated eggs. red blood cells, antisera	SOP 11.05, based on ANON (2012) Avian Influenza: Manual of diagnostic tests and vaccines for terrestrial animals. Chapter 2.3.14, (2012) OIE., 12.1. Anon (1992). Council Directive 92/66/EEC of 14th July 1992 introducing community measures for the control of Newcastle Disease. Official Journal of the European Communities L260, pg 1-20 Annex III	
815 Detection of prions using specific antibodies and appropriate techniques - .01 Histological identification of prion disease lesions	Detection of Disease specific Prion protein by Bio-Rad TeSeE Western Blot	SOP 2.14, SOP 3.15 based on TeSeE Western Blot.	tissue	Bio-Rad Western Blot Assay Kit	OIE approved registration number 20090105	
	Haematoxylin & Eosin staining of suspect TSE samples	SOP 2.4, SOP 3.16	formalin fixed tissue	Leica stainer & coverslipper	SOP 2.4, SOP 3.16 based on OIE Terrestrial Manual chapters 2.4.6 Ed 2010 (BSE), 2.7.13 Ed 2009 (Scrapie)	
815 Detection of prions using specific antibodies and appropriate techniques - .02 Detection of prion protein by immunological methods (including ELISA, Western Blots, immunohistochemistry)	Detection of Disease specific Prion protein by Bio-Rad TeSeE based Hybrid Western Blot	SOP 2.63	tissue	Bio-Rad Western Blot Assay kit	SOP 2.63, based on TeSeE based Hybrid Western Blot. Approved by EURL strain-typing expert group	
	Immunohistochemistry of suspect TSE samples	SOP 2.1, SOP 2.2, SOP 3.16	tissue	Ventana Immunohistochemistry automated stainer	SOP 2.1, SOP 2.2, SOP 3.16 based on OIE Terrestrial Manual chapters 2.4.6 Ed 2010	

					(BSE), 2.7.13 Ed 2009 (Scrapie)	
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